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HYDROCARBON RECEPTOR ANTAGONISTIC LIGANDS
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TRANSMITTAL LETTER

Enclosed herewith for filing in connection with the above-identified patents please find the following:

1. A Certified Copy of Priority Document, European Application No. EP 02 292 786.7; and
2. Return Postcard.

In connection with the foregoing matters, please charge any additional fees which may be due, or credit any overpayment, to Deposit Account Number 50-1133.

Respectfully submitted,

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Die angehefteten Unterlagen stimmen mit der ursprünglich eingereichten Fassung der auf dem nächsten Blatt bezeichneten europäischen Patentanmeldung überein.

The attached documents are exact copies of the European patent application described on the following page, as originally filed.

Les documents fixés à cette attestation sont conformes à la version initialement déposée de la demande de brevet européen spécifiée à la page suivante.

Patentanmeldung Nr. Patent application No. Demande de brevet n°

02292786.7

Der Präsident des Europäischen Patentamt:
Im Auftrag

For the President of the European Patent O

Le Président de l'Office européen des brev
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R C van Dijk

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Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.
If no title is shown please refer to the description.
Si aucun titre n'est indiqué se référer à la description.)

New stilbene derivatives and their use as aryl hydrocarbon receptor antagonistic
ligands

In Anspruch genommene Priorität(en) / Priority(ies) claimed /Priorité(s)
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**"NEW STILBENE DERIVATIVES AND THEIR USE AS ARYL HYDROCARBON
RECEPTOR ANTAGONISTIC LIGANDS"**

The invention relates to new stilbene derivatives having, particularly, specific antagonistic ligand properties with respect to the Aryl Hydrocarbon Receptor (AhR) and their use for the prevention and treatment of poisoning and pathologies caused by toxic aryl hydrocarbons and other ligands of the AhR.

5

Halogenated (HAH) and polycyclic (PAH) aryl hydrocarbons, polyaromatic hydrocarbons, polychlorinated biphenyls (PCB) and other industrial chemicals that bind to the aryl hydrocarbon receptor (AhR) are environmental contaminants which will be collectively referred to hereinafter by the term AhR ligands.

10

AhR ligands have attracted much attention and concern recently because of their resistance to degradation, resulting in a long biologic half-life (> 10 years) in soil (see ref. 1) and between 4 and 12 years in human blood and fat (2).

15 Every person on earth is continually exposed to AhR ligands which are present in cigarette smoke, in exhaust fumes from both gasoline and diesel engines, in furnace gases, in cooked meat and fish, in dairy products, and even in mother's milk.

20 There is sufficient evidence to link exposure to AhR ligands to the development of numerous pathologies such as atherosclerosis, cancer, immunosuppression, skin disorders, reproductive failure, and diminished resistance to viral infection.

25 Attempts to reduce exposure to said environmental toxins have not been successful, nor are they likely to be until internal combustion engines, the use of fossil fuels, and cigarette smoking are eliminated.

It is, therefore, imperative to develop methods to antagonize the adverse effects of toxic AhR ligands. In WO 99/56737 in the name of INSERM, co-inventors of the present patent application disclosed the potent anti-dioxin effects of resveratrol and analogs and gave a valid
30 response to this challenge.

The inventors have elaborated new stilbene derivatives and have surprisingly found more active and more selective AhR antagonists than resveratrol.

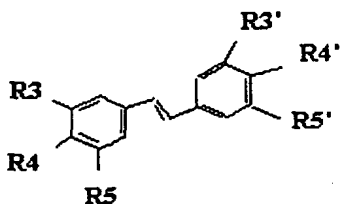
5 Advantageously contrary to resveratrol, such derivatives are devoid of affinity for the estrogen receptor (ER).

Accordingly, the primary object of the invention is to provide new stilbene derivatives with antagonist properties with respect to AhR ligands.

10

Another object of the invention is to provide drugs for preventing or treating pathologies and poisoning induced by exposure to AhR ligands.

The stilbene derivatives of the invention have formula I :



15

wherein

20 R3, R4 and R5 and R3', R4' and R5' are identical or different and represent H, OH, O-alkoxy or hal, said alkoxy group being a C1-C6 alkoxy and "hal" being F, Cl or CF₃, with the proviso that one of R4', R3 and R5 or R4, R3' and R5' does not represent OH, OCH₃, or OCH₂ CH₃ when the two other substituents are OH, OCH₃, or OCH₂ CH₃, respectively,

and the symmetrical derivatives.

25

As shown by the results given in the Examples, said derivatives have a high affinity for the AhR, but without detectable affinity for the ER.

Said derivatives are in racemic form or in the form of geometric cis or trans isomers.

30

Preferred stilbene derivatives are trans isomers.

In a preferred group, R3 and R5 are hal.

- 5 In advantageous derivatives of said group R3' or R4' is hal, alkoxy or hydroxy, and R5' is H.

Preferably R3 and R5 are Cl and in particularly preferred derivatives R3' is H and R4' or R5' is methoxy or Cl.

- 10 The invention particularly relates to (E)-1-(4'-trifluoromethylphenyl)-2-(3,5-ditrifluoromethylphenyl)-ethene, (E)-1-(4'-methoxyphenyl)-2-(3,5-dichlorophenyl)-ethene, and (E)-1-(4'-chlorophenyl)-2-(3,5-dichlorophenyl)-ethene which bound to AhR with respective relative binding affinity (RBA) of 52.1, 112.0 and 130.0 without detectable affinity for ER.

15

The invention also relates to the symmetrical compounds, the positions 4'-3,5 and 4-3',5' with respect to said substituents being identical. The invention particularly relates to the derivatives wherein R3',R5' and R4 represent hal or R3' and R5' are hal and R4 is OH or alkoxy.

20

Advantageously, said compounds are not toxic on cell culture up to 10 μ M .

They are particularly useful to investigate further the role of AhR both at the physiological level and in pathologies in which this receptor is involved through activation by xenobiotics.

25

Said derivatives are useful as antagonists to AhR ligands for binding to the AhR receptor.

- 30 Their absence of toxicity make them of interest as active principle in therapeutic and nutritional fields, and in general for the prevention or treatment of disorders due to the toxic effects resulting from exposure to AhR ligands.

The pharmaceutical compositions of the invention comprise an effective amount of at least one stilbene derivative as above defined with pharmaceutically acceptable carrier.

Said pharmaceutical compositions are advantageously in a form for administration by the oral, nasal, parenteral or topical route.

5 The drugs thus developed are presented in forms appropriate for administration by the oral route, such as drops, gel capsules, syrup or alcohol syrup, by the nasal route, such as spray or drops, by the parenteral route, in the form of a solution in an appropriate solvent, or again by the topical route, such as cream, ointment, shampoo or lotion. The pharmaceutically acceptable vehicles or excipients used are based on oils authorized in the pharmacopoeia, or on an alcohol such as ethanol or propylene-glycol (from 10% to 100%) or on DMSO. In
10 general, they are non-aqueous solvents. Use can also be made of a lipid preparation such as Intralipid^R.

It will be noted that said antagonistic stilbenes are soluble in the excipients and/or vehicles used for the preparation of drugs.

15

The dosage in the different forms and for daily administration will be established, in the standard way, depending on the type of effects to be treated. By way of example, the said drugs based on the said antagonists will be administered in an amount of from 0.1 mg to 500 mg/day, especially from 20 mg to 200 mg/day, and in particular from 10 to 100 mg/day.

20

Higher doses of stilbene derivatives, up to 5 g per day, can be administered as an antidote for acute intoxication by AhR ligands.

25 The drugs developed on the basis of these antagonists can be used in various pathologies involving AhR ligands, such as atherogenesis, immunosuppression, cancer, viral infections such as AIDS, allergies and dermatological diseases associated with dioxin and the AhR ligands and with osteoporosis.

30 The inhibitory effects of the said stilbenes towards the AhR ligands are also advantageously exploited in dietetics.

The invention also concerns the use of the above-defined antagonists as food additives.

Thus it concerns foods characterized in that they contain at least one antagonist such as defined above in a quantity allowing it to exert an inhibitory effect towards AhR ligands, so as to prevent their harmful effects. These foods are intended for adults or for children and infants.

5

These additives are incorporated in the food, for example in powdered milk, liquid or solid preparations (cereals) or canned foods.

They can also be dissolved in oil for administration as drops or as a food additive.

10

They can be incorporated during the production of the foods or when they are packaged, or by using preparations with the antagonists already formulated, making it possible to effect the desired supplementation.

15 The dose used will be reduced compared to that adopted for an adult, taking account of the weight of the child and its food, so as to obtain an effective plasma concentration (of the order of micromolar).

20 The antagonists of the invention are likewise of great value for combating the effects of smoking, which corresponds to chronic exposure to AhR ligands.

25 The advantageous properties of the antagonists of the invention are exploited by using them to impregnate cigarette filters, so as to deliver a dose of stilbene derivatives proportional to the concentration of the poisons absorbed with the smoke, and leading to a blood concentration making it possible to inhibit toxic effects of AhR ligands.

Other characteristics and advantages of the invention are given in the examples that follow, which, by way of illustration.

30 Experimental Section

Chemicals : 2,3,7,8-tetrachloro (1,6-3H) dibenzo-p-dioxin, 28 Ci/mmol was purchased from Terrachem (Lenexa, KS). Dioxin stock solutions were initially dissolved in dimethylsulfoxide and handled under a fume hood. TCDD stock was subsequently diluted in ethanol for use in

experiments described below. Steroids were purchased from Steraloids (Wilton, NH). All other chemicals were purchased from Sigma Chemicals.

Chemistry

5 ¹H NMR were recorded at 200 MHz on a Bruker AC200 spectrometer. Chemical shifts are given in parts per million and tetramethylsilane was used as the internal standard for spectra obtained in acetone-d₆. All J values are given in Hz. Mass spectra were recorded on a variant MAT 311 A mass spectrometer. Elemental analysis was carried out by the microanalytical service laboratory of the Ecole Supérieure de Chimie of the Université Paul Sabatier, France
10 and all values were within ±0.4% of the calculated composition. Reagents and solvents were used as obtained from commercial suppliers without further purification. Reaction progress was determined by TLC analysis on silica gel coated aluminium plates. Visualization with UV light (254 nm). All stilbene were purified by RP HPLC and detected by UV light (254 nm). Melting points were uncorrected and measured on a Kofler apparatus. Phosphonium
15 chloride **2a-g** were prepared by refluxing a stirred mixture of triphenylphosphine and the corresponding benzyl chloride in toluene. Yields were better than 50% for all experiments.

General procedure for the preparation of stilbenes (**Z**) and stilbenes (**E**)

Phosphonium salt (1mmol) was dissolved in dichloromethane(1ml). Aryl aldehyde (1mmol),
20 18 crown 6 (0,1 mmol) and potassium hydroxyde (3mmol) was added to the solution. Reaction mixture was stirred at room temperature and periodically monitored by TLC (toluene or hexane) until complete consumption of the aldehyde. The mixture was dilute with dichloromethane and filtered. The organic layer was washed with, dried over MgSO₄ and evaporated under vacuum. Purification by RP HPLC (Ultrasep ES 100 RP 18, 250 x 8 mm,
25 6.0 µm; methanol :water, 80:20 or 85:15 ; flow rate 1ml/min) afford pure product.

(E)-1-(4-methoxyphenyl)-2-(3,5-dimethoxyphenyl)-ethene (4a)

Mp : 55-57°C ; Rf (toluene) : 0,4 ; ^1H NMR (acetone- d_6) : 3.808 (s, 6H), 3.813 (s, 3H), 6.387 (t, $J=2.25\text{Hz}$, 1H), 6.743 (d, $J=2.5\text{Hz}$, 2H), 6.94 (m, 2H), 7.05 (d, $J=16.5\text{Hz}$, 1H), 7.17 (d, $J=16.5\text{Hz}$, 1H), 7.53 (m, 2H); ^{13}C NMR (acetone- d_6): 55.575, 100.245, 105.056, 114.957, 127.285, 128.661, 129.463, 130.876, 140.746, 160.489, 162.081; MS (DCI/ NH_3) : m/z 271 (M+1); UV : $\lambda_1 = 307\text{ nm}$ $\epsilon = 24\,600\text{ M}^{-1}\cdot\text{cm}^{-1}$, $\lambda_2 = 317\text{ nm}$ $\epsilon = 23\,800\text{ M}^{-1}\cdot\text{cm}^{-1}$

(E)-1-(4-chlorophenyl)-2-(3,5-dichlorophenyl)-ethene (4b)

Mp: 94-96°C ; Rf (hexane): 0.56; ^1H NMR (acetone- d_6): 7.257 (d, 1H, $J = 16.44\text{Hz}$, 1H), 7.369 (t, $J = 1.9\text{ Hz}$, 1H), 7.4-7.49 (m, 2H), 7.448 (d, 1H, $J = 16.32\text{Hz}$, 1H), 7.62 (d, $J = 1.9\text{ Hz}$, 2H), 7.625-7.68 (m, 2H); ^{13}C NMR (acetone- d_6): 125.84, 127.361, 127.77, 129.292, 129.727, 135.872; MS (DCI/ CH_4): m/z : 247 ($\text{MH}^+ - \text{HCl}$; 47), 283 (MH^+ ; 100), 311 ($\text{M} \dots \text{C}_2\text{H}_5^+$; 35), 323 ($\text{M} \dots \text{C}_3\text{H}_5^+$; 2.2); UV : $\lambda_1 = 301\text{ nm}$ $\epsilon = 20\,500\text{ M}^{-1}\cdot\text{cm}^{-1}$, $\lambda_2 = 313\text{ nm}$ $\epsilon = 20\,500\text{ M}^{-1}\cdot\text{cm}^{-1}$.

(E)-1-(4-methoxyphenyl)-2-(3,5-difluorophenyl)-ethene (4c)

Mp: 108-110°C; Rf (toluene 1 / hexane 1) : 0.58; ^1H NMR (acetone- d_6): 3.826 (s, 3H), 6.85 (tt, 1H, $J_1 = 10.1\text{ Hz}$, $J_2 = 2.3\text{ Hz}$, 1H), 6.96 (m, 2H), 7.57 (m, 2H), 7.09 (d, $J = 16.4\text{ Hz}$, 1H), 7.34 (d, $J = 16.4\text{ Hz}$, 1H), 7.21 (dt, $J_1 = 7.14\text{ Hz}$, $J_2 = 1.6\text{ Hz}$, 2H); ^{13}C NMR (acetone- d_6): 55.619, 102.607, 109.56, 115.052, 124.834, 129.115, 130.119, 132.122, 142.894, 161.011, 164.23; MS (DCI/ CH_4): $m/z = 227$ ($\text{MH}^+ - \text{HF}$; 15.8), 246 (M^+ ; 100), 275 ($\text{M} \dots \text{C}_2\text{H}_5^+$; 13.5); UV : $\lambda_1 = 308\text{ nm}$ $\epsilon = 46\,320\text{ M}^{-1}\cdot\text{cm}^{-1}$, $\lambda_2 = 320\text{ nm}$ $\epsilon = 49\,800\text{ M}^{-1}\cdot\text{cm}^{-1}$.

(E)-1-(4-fluorophenyl)-2-(3,5-difluorophenyl)-ethene (4d)

Mp : 47 °C ; Rf (hexane) : 0.34; ^1H NMR (acetone- d_6): 6.91 (tt, $J_1 = 9.5\text{ Hz}$, $J_2 = 2.2\text{Hz}$, 1H), 6.743 (d, 2H, $J = 2.5\text{Hz}$, 2H), 7.17-7.28 (m, 5H), 7.41 (d, $J = 16.42\text{Hz}$, 1H), 7.68 (m, 2H); ^{13}C NMR (acetone- d_6): 103.1, 109.9, 116.445, 127.123, 129.63, 131.23, 142.364, 163.56, 164.2 MS (DCI/ CH_4): $m/z = 215$ ($\text{MH}^+ - \text{HF}$; 38), 234 (M^+ ; 100), 263 ($\text{M} \dots \text{C}_2\text{H}_5^+$; 23.47), 275 ($\text{M} \dots \text{C}_3\text{H}_5^+$; 1.04); UV : $\lambda_1 = 295\text{ nm}$ $\epsilon = 23\,900\text{ M}^{-1}\cdot\text{cm}^{-1}$, $\lambda_2 = 307\text{ nm}$ $\epsilon = 22\,100\text{ M}^{-1}\cdot\text{cm}^{-1}$

(E)-1-(4-trifluoromethylphenyl)-2-(3,5-difluoromethylphenyl)-ethene (4e)

Mp :99-100°C ;Rf (hexane) : 0.39 ; ^1H NMR (acetone- d_6): 7.65 (d, $J=16.6$ Hz, 1H), 7.77 (d, $J=16.6$ Hz, 1H), 7.76 (m, 2H), 7.91 (m, 2H), 7.96 (s,1H), 8.31 (s,2H) ; ^{13}C NMR (acetone d_6) 121.870, 126.541, 127.857, 128.371, 129.136, 132.051; MS (DCI/ CH_4): m/z =
 5 365(MH^+ -HF ;100), 385 (MH^+ ; 23.58), 413 ($\text{M} \dots \text{C}_2\text{H}_5^+$; 12.5), 425 ($\text{M} \dots \text{C}_3\text{H}_5^+$; 1.95); UV :
 $\lambda_1 = 297$ nm $\epsilon = 28\,200 \text{ M}^{-1} \cdot \text{cm}^{-1}$, $\lambda_2 = 307$ nm $\epsilon = 25\,800 \text{ M}^{-1} \cdot \text{cm}^{-1}$

(E)-1-(4-fluorophenyl)-2-(3,5-dimethoxyphenyl)-ethene (4f)

Mp: 46-48°C; Rf (toluene): 0.53; ^1H NMR (acetone- d_6): 3.81 (s, 6H), 6.42 (t, $J=2.25$ Hz, 1H), 6.77 (d, $J=2.26$ Hz, 2H), 7.08-7.182 (m, 3H), 7.269 (d, $J=16.4$ Hz, 1H), 7.635 (m, 2H);
 10 ^{13}C NMR: 55.601, 100.666, 105.347, 116.28, 128.591, 129.172, 129.522; MS (DCI/ NH_3):
 $m/z = 259$ (MH^+ ;100) ;UV : $\lambda_1 = 297$ nm $\epsilon = 22\,900 \text{ M}^{-1} \cdot \text{cm}^{-1}$, $\lambda_2 = 308$ nm $\epsilon = 21\,200 \text{ M}^{-1} \cdot \text{cm}^{-1}$

(E)-1-(4-ethoxyphenyl)-2-(3,5-dimethoxyphenyl)-ethene (4g)

Mp: 54-55°C; Rf (toluene): 0.45; ^1H NMR (acetone- d_6): 1.366 (t, $J=6.955$ Hz, 3H), 3.808(s, 6H), 4.06 (q, $J=6.98$ Hz, 2H), 6.385 (t, $J=2.25$ Hz, 1H), 6.741 (d, $J=2.23$ Hz, 2H),
 15 6.44(d, $J=16.1$ Hz, 1H), 6.55 (d, $J=16.1$ Hz, 1H), 6.918(m, 2H), 7.513(m, 2H); ^{13}C NMR
 (acetone- d_6): 15.094, 55.564, 64.012, 100.196, 105.022 , 115.441, 127.157, 128.661, 129.492,
 130.725, 140.757, 159.812, 162.07; MS (DCI/ NH_3): m/z 285 ($\text{M}+1$, 100); UV.: $\lambda_1 = 307$ nm
 $\epsilon = 36\,900 \text{ M}^{-1} \cdot \text{cm}^{-1}$, $\lambda_2 = 320$ nm $\epsilon = 36\,800 \text{ M}^{-1} \cdot \text{cm}^{-1}$

(E)-1-(4-butoxyphenyl)-2-(3,5-dimethoxyphenyl)-ethene (4h)

20 Mp: 54-56°C; Rf (toluene): 0.51; ^1H NMR (acetone- d_6): 0.9666 (t, $J=7.28$ Hz, 3H), 1.49
 (m,2H) 1.755 (m,2H), 3.808 (s,6H), 4.007 (t, $J=6.35$ Hz ,2H), 6.3866 (t, $J=2.24$ Hz, 1H),
 6.742 (d, $J=2.25$ Hz, 2H), 6.927 (m, 2H), 7.512 (m, 2H) 7.016 (d, $J=16.36$ Hz, 1H), 7.2 (d,
 $J=16.36$ Hz, 1H); ^{13}C NMR (acetone- d_6): 14.109, 19.888, 32.076, 55.566, 68.256, 100.206,
 105.035, 115.485, 127.148, 128.653, 129.505, 130.712, 140.76, 159.995, 162.075 ; MS

(DCI/NH₃): m/z 313 ($M+1$, 100); UV : $\lambda_1 = 307$ nm $\epsilon = 40\,100$ M⁻¹.cm⁻¹, $\lambda_2 = 320$ nm $\epsilon = 40\,000$ M⁻¹.cm⁻¹.

(E)-1-(4-trifluoromethylphenyl)-2-(3,5-dichlorophenyl)-ethene (4i)

Mp: 98-100°C; R_f (hexane): 0.43; ¹H NMR (acetone-*d*₆): 7.395 (d, $J = 16.54$ Hz, 1H), 7.561

5 (d, $J = 16.54$ Hz, 1H), 7.4107 (t, $J = 1.87$ Hz, 1H), 7.674 (d, $J = 1.84$ Hz, 2H), 7.74 (m, 2H), 7.86

(m, 2H); ¹³C NMR (acetone-*d*₆): 126.116, 126.569, 128.223, 129.352, 131.098, 135.93

MS (DCI/CH₄): m/z 281 ($MH^+ - HCl$; 11.8), 297 ($MH^+ - HF$; 100), 317 (MH^+ ; 79.7), 345

($M \dots C_2H_5^+$; 24.8); UV : $\lambda_1 = 297$ nm $\epsilon = 41\,000$ M⁻¹.cm⁻¹, $\lambda_2 = 309$ nm $\epsilon = 41\,100$ M⁻¹.cm⁻¹

, $\lambda_3 = 324$ nm $\epsilon = 25\,000$ M⁻¹.cm⁻¹

10 **(E)-1-(4-methoxyphenyl)-2-(3,5-dichlorophenyl)-ethene (4j)**

Mp: 62-64°C; R_f (toluene/hexane): 0.7; ¹H NMR (acetone-*d*₆): 3.828 (s, 3H), 6.96 (m, 2H),

7.07); ¹³C NMR (acetone-*d*₆): 55.621, 115.045, 124.118, 125.382, 127.011, 129.161,

132.419; MS (DCI/NH₃): m/z 278 (MH^+ ; 100), 296 ($MH^+ \dots NH_3$; 15.4); UV : $\lambda_1 = 309$ nm $\epsilon =$

44 400 M⁻¹.cm⁻¹, $\lambda_2 = 325$ nm $\epsilon = 51\,600$ M⁻¹.cm⁻¹

15 **(Z)-1-(4-methoxyphenyl)-2-(3,5-dimethoxyphenyl)-ethene (5a)**

colourless oil; R_f (toluene) : 0.4; ¹H NMR (acetone-*d*₆): 3.66 (s, 6H), 3.77 (s, 3H), 6.34 (t,

$J = 2.16$ Hz, 1H), 6.429 (d, $J = 2.7$ Hz, 2H), 6.45 (d, $J = 12.4$ Hz, 1H), 6.556 (d, $J = 12.4$ Hz, 1H),

6.824 (m, 2H), 7.218 (m, 2H); ¹³C NMR (acetone-*d*₆): 55.418, 55.47, 100.085, 107.372,

114.387, 129.443, 130.315, 130.883, 131.014, 140.307, 159.925, 161.706; MS (DCI/NH₃):

20 m/z 271 (MH^+); UV : $\lambda_{max} = 286$ nm $\epsilon = 25\,500$ M⁻¹.cm⁻¹

(Z)-1-(4-chlorophenyl)-2-(3,5-dichlorophenyl)-ethene (5b)

colourless oil; R_f (hexane): 0.63; ¹H NMR (acetone-*d*₆): 6.63 (d, $J = 12.18$ Hz, 1H) 6.77 (d,

$J = 12.18$ Hz, 1H), 7.19 (d, $J = 1.9$ Hz, 2H), 7.23-7.36 (m, 5H); ¹³C NMR (acetone-*d*₆):

127.748, 128.087, 128.932, 129.467, 131.327, 132.398, 133.899, 135.494, 135.87, 141.375

MS (DCI/CH₄): m/z : 283 (MH⁺; 100), 311 (M...C₂H₅⁺; 46), 323 (M...C₃H₅⁺; 3.6); UV : λ_{\max}
= 286 nm ϵ = 13 600 M⁻¹.cm⁻¹

(Z)-1-(4-methoxyphenyl)-2-(3,5-difluorophenyl)-ethene (5c)

colourless oil; R_f (toluene 1 / hexane 1) : 0.62; ¹H NMR (acetone-*d*₆): 3.788 (s, 3H), 6.497 (d, J = 12.15 Hz, 1H), 6.706 (d, J = 12.15 Hz, 1H), 6.819-6.88 (m, 5H), 7.19(m, 2H); ¹³C NMR (acetone-*d*₆): 55,506, 102.915, 112.3, 114.678, 127.067, 130.97, 133.103; MS (DCI/CH₄): m/z =227 (MH⁺-HF;30.37), 247 (MH⁺;100), 275 (M...C₂H₅⁺; 18.28), 287 (M...C₃H₅⁺); UV : λ_{\max} = 291 nm ϵ = 14 000 M⁻¹.cm⁻¹

(Z)-1-(4-fluorophenyl)-2-(3,5-difluorophenyl)-ethene (5d)

colourless oil; R_f (hexane) : 0.41; ¹H NMR (acetone-*d*₆): 6.62 (d, J= 12.2Hz, 1H), 6.81- 6.9 (m,4H), 7.25-7.32(m, 2H); ¹³C NMR (acetone-*d*₆): 103.2, 112.4, 116.16, 128.86, 131.63,132.3; MS (DCI/CH₄): m/z = 215 (MH⁺-HF; 17), 234 (M⁺;100), 263 (M..C₂H₅⁺; 7), 275 (M...C₃H₅⁺; 0.45); UV : λ_{\max} = 277 nm ϵ = 6000 M⁻¹.cm⁻¹

(Z)-1-(4-trifluoromethylphenyl)-2-(3,5-difluoromethylphenyl)-ethene (5e)

colourless oil; R_f (hexane) : 0.39; ¹H NMR (acetone-*d*₆): 6.97 (d, J=12.2 Hz, 1H), 7.05 (d, J=12.2 Hz, 1H), 7.48 (m, 2H), 7.66 (m, 2H), 7.797 (s, 2H), 7.89 (s, 1H); ¹³C NMR (acetone-*d*₆): 121.697, 126.403, 130.089, 130.277, 133.211; MS (DCI/CH₄): m/z = 365 (MH⁺-HF ;100), 385 (MH⁺ ;17.9), 413 (M...C₂H₅⁺; 15.5), 425 (M...C₃H₅⁺; 3.7); UV : λ_{\max} = 276 nm ϵ = 9000 M⁻¹.cm⁻¹

(Z)-1-(4-fluorophenyl)-2-(3,5-dimethoxyphenyl)-ethene (5f)

colourless oil; R_f (toluene): 0.59; ¹H NMR (acetone-*d*₆): 3.657 (s, 6H), 6.362 (t, J =2.25 Hz, 1H), 6.39 (d, J =2.2 Hz, 2H), 6.556 (d, J = 12.2 Hz, 1H), 6.625 (d, J = 12.2 Hz, 1H), 7.037 (m, 2H), 7.306 (m, 2H); ¹³C NMR (acetone-*d*₆): 55.425, 100.378, 107.421, 115.81, 130.068, 131.194, 131.647 ; MS (DCI/NH₃): m/z = 259 (MH⁺ ; 100); UV : λ_{\max} = 277 nm ϵ = 39 000 M⁻¹.cm⁻¹

(Z)-1-(4-ethoxyphenyl)-2-(3,5-dimethoxyphenyl)-ethene (5g)

colourless oil; R_f (toluene): 0.5; ¹H NMR (acetone-*d*₆): 1.34 (t, J = 6.96 Hz, 3H), 3.66(s,6H), 4.02 (q, J = 6.98 Hz, 2H), 6.337 (t, J = 2.27 Hz, 1H), 6.428 (d, J = 2.28 Hz, 2H), 6.44 (d, J = 12.28 Hz, 1H), 6.55 (d, J = 12.28 Hz, 1H), 7.192(m, 2H), 7.225 (m, 2H); ¹³C NMR (acetone-*d*₆): 15.059, 55.414, 63.913, 100.073, 107.357, 114.886, 129.355, 130.75, 130.926, 131.02, 140.321, 161.703; MS(DCI/NH₃): *m/z* 285(M+1, 100); 302(MH⁺....NH₃); UV : λ_{max} = 285 nm ε = 24 000 M⁻¹.cm⁻¹

(Z)-1-(4-butoxyphenyl)-2-(3,5-dimethoxyphenyl)-ethene (5h)

colourless oil; R_f (toluene): 0.56; ¹H NMR (acetone-*d*₆): 0.952 (t, J = 7.27 Hz, 3H), 1.47 (m, 2H), 1.688-1.76 (m, 2H), 3.662 (s, 6H), 3.968 (t, J = 7.1 Hz, 2H), 6.339 (t, J = 2.28 Hz, 1H), 6.435 (d, J = 2.68 Hz, 2H), 6.44 (d, J = 12.3 Hz, 1H), 6.55 (d, J = 12.3 Hz, 1H), 6.815 (m, 2H), 7.209 (m, 2H); ¹³C NMR (acetone-*d*₆): 14.103, 19.866, 32.041, 55.413, 68.152, 100.053, 107.357, 114.918, 129.338, 130.154, 130.932, 131.013, 140.329, 159.416, 161.691 MS (DCI/NH₃): *m/z* 313 (M+1, 100); UV : λ_{max} = 286 nm ε = 38 200 M⁻¹.cm⁻¹

(Z)-1-(4-trifluoromethylphenyl)-2-(3,5-dichlorophenyl)-ethene (5i)

colourless oil; R_f (hexane): 0.48; ¹H NMR (acetone-*d*₆): 6.764 (d, J = 12.52 Hz, 1H), 6.891 (d, J = 12.52 Hz, 1H), 7.195 (d, J = 1.9 Hz, 2H), 7.3646 (t, J = 1.93 Hz, 1H), 7.47 (m, 2H), 7.66 (m, 2H); ¹³C NMR (acetone-*d*₆): 126.216, 127.962, 128.147, 130.348, 132.251 MS (DCI/CH₄): *m/z* 281 (MH⁺-HCl; 8.5), 297 (MH⁺-HF; 100), 317 (MH⁺; 68.4), 345 (M....C₂H₅⁺; 28.96); UV : λ_{max} = 276 nm ε = 38 900 M⁻¹.cm⁻¹

(Z)-1-(4-methoxyphenyl)-2-(3,5-dichlorophenyl)-ethene (5j)

colourless oil; R_f (toluene/hexane): 0.75; ¹H NMR (acetone-*d*₆): 3.79 (s, 3H), 6.476 (d, J = 12.12 Hz, 1H), 6.716 (d, J = 12.12 Hz, 1H), 6.857 (m, 2H), 7.2 (m, 4H), 7.313 (t, J = 1.93 Hz, 1H); ¹³C NMR (acetone-*d*₆): 55.529, 114.7, 126.407, 127.332, 128.026, 133.384, 135.353

MS(DCI/NH₃): *m/z* 278 (MH⁺;100), 296 (MH⁺....NH₃; 16.24); UV : λ_{max} = 296 nm ϵ = 12
300 M⁻¹.cm⁻¹

AhR and ER-mediated transactivation.

Cis and Trans isomers were then tested for transcriptional modulatory activity in a DRE-TK-
5 CAT containing stable cell line treated with TCDD. All the compounds displayed antagonist
activities on AhR.

To assay the β estrogenic properties of resveratrol, tests were performed with a stable cell
line expressing an ERE-driven Luciferase reporter gene for ER-mediated transcriptional
regulation. Results showed that the presence of oxygen atom on substituents yielded
compounds with weak agonistic activities with regard to estradiol through ER. None of the
other compounds displayed any antagonistic activities through ER. Results for both lines of
experiments are summarized on Tables I and II.

Table I: Characteristics of *trans*-Stilbenes derivatives.

mp°C: melting point. Ago: agonist. N.M.: not measurable. RBA: relative binding affinity of
the compound for the given receptor (AhR or ER). The estimated in vitro affinity of
resveratrol for the AhR (calculated as IC₅₀) was 6.10⁻⁶ M. R.T.A.: Residual Transcriptional
Activity: This is calculated as the residual induction of the stably transfected DRE-TKCAT
reporter gene when 10⁻⁹ M of TCDD is challenged by 10⁻⁷ M of the compound, expressed as
average \pm SEM. (n = 3 to 6 determinations).

Compound s	R ₁	R ₂	mp °C	AhR		ER	
				RBA	T.A.	RBA	T.A.
Resveratrol	4'-OH	3,5-OH		1	Ant	0.01	Ago
4a	4'-OMe	3,5-OMe	55-57	21.0	Ant	0.03	Ago
4b	4'-Cl	3,5-Cl	94-96	130.0	Ant	N.M.	N.M.
4c	4'-OMe	3,5-F	108-110	17.5	Ant	N.M.	N.M.
4d	4'-F	3,5-F	47-48	39.1	Ant	N.M.	N.M.
4e	4'-CF ₃	3,5-CF ₃	99-100	52.1	Ant	N.M.	N.M.
4f	4'-F	3,5-OMe	46-48	32.1	Ant	N.M.	N.M.
4g	3',5'-OMe	4-OEt	54-55	39.8	Ant	N.M.	N.M.
4h	3',5'-OMe	4-OBu	54-56	8.1	Ant	N.M.	N.M.
4i	4'-CF ₃	3,5-Cl	98-100	791.0	Ago	N.M.	N.M.
4j	4'-OMe	3,5-Cl	62-64	112.0	Ant	N.M.	N.M.
4k	3'-CF ₃	3,5-Cl	110-111	26.3	Ant	N.M.	N.M.

Table II: Characteristics of *cis*-Stilbenes derivatives.

mp°C: melting point. Ago: agonist. N.M.: not measurable. RBA: relative binding affinity of the compound for the given receptor (AhR or ER). The estimated in vitro affinity of resveratrol for the AhR (calculated as IC₅₀) was 6.10⁻⁶ M. R.T.A.: Residual Transcriptional Activity: This is calculated as the residual induction of the stably transfected DRE-TKCAT reporter gene when 10⁻⁹ M of TCDD is challenged by 10⁻⁷ M of the compound, expressed as average \pm SEM. (n = 3 to 6 determinations).

Compounds	R ₁	R ₂	mp °C	AhR		ER	
				RBA	TA	RBA	TA
5a	4'-OMe	3,5-OMe	oil	2.1	Ant	0.06	Ago
5b	4'-Cl	3,5-Cl	oil	12.4	Ant	N.M.	N.M.
5c	4'-OMe	3,5-F	oil	7.2	Ant	N.M.	N.M.
5d	4'-F	3,5-F	oil	2.6	Ant	N.M.	N.M.
5e	4'-CF ₃	3,5-CF ₃	oil	2.7	Ant	N.M.	N.M.
5f	4'-F	3,5-OMe	oil	1.7	Ant	N.M.	N.M.
5g	3',5'-OMe	4-OEt	oil	2.5	Ant	N.M.	N.M.
5h	3',5'-OMe	4-OBu	oil	3.7	Ant	N.M.	N.M.
5i	4'-CF ₃	3,5-Cl	oil	10.9	Ant	N.M.	N.M.
5j	4'-OMe	3,5-Cl	oil	13.0	Ant	N.M.	N.M.
5k	4'-OMe	3,5-CF ₃	oil	6.3	Ant	N.M.	N.M.
5l	3'-OMe	3,5-Cl	oil	7.1	Ant	N.M.	N.M.

Measurements of cytotoxicity

Cytotoxicity was evaluated of the A549 cell line. Resveratrol and its halogenated homologs

10 were not toxic up to a 10 microM concentration.

Cell culture and gene reporter assays.

5 The human adenocarcinoma breast cell line MCF-7 was obtained from the American Tissue Culture Collection (Rockville, USA). The cells were established by stably transfecting MCF-7 cells with the ERE-globin-tk-luc-SV-Neo plasmid and thus expressed luciferase in an estrogen dependent manner MCF-7-Luc. MCF-7 cells were grown routinely in RPMI 1640 medium and (MCF-7-Luc) cells in DMEM medium, supplemented with 5% FBS (Gibco
10 BRL, Life Technologies, Cergy Pontoise, France). Cells were incubated at 37°C in a humidified 5% CO₂ incubator. For experiments, cells were grown for 5 days in phenol red-free medium, containing 5% dextran-coated charcoal treated FCS (DCC-FCS). Medium was changed after 2 days. At day 5, cells were treated or not with compounds. Compounds were dissolved in ethanol.

15 Luciferase assay:

For each condition, 15.103 cells were seeded per well in 12-well plates and treated, as described above, during 16 hours in a final volume of 0.5 mL. At the end of the treatment, cells were washed with PBS and lysed in 150 µl of lysis buffer (Promega, Charbonnières, France). Luciferase activity was measured using the luciferase assay reagent (Promega),
20 according to the manufacturer's instructions. Protein concentrations were measured using Bradford technique, to normalize the luciferase activity data. For each condition, average luciferase activity was calculated from the data of 3 independent wells.

The stable cell line 47DRE bearing the TCDD-responsive CAT construct was previously
25 described (3). It was routinely grown in DMEM medium supplemented with 10 % fetal calf serum (FCS), 4,5 g/L glucose, and 0.6 units/ml (10⁻⁶ M) insulin. Unless stated otherwise, cells were established 24 hours before any experiment in a modified medium (stripped condition) containing 5% Dextran-charcoal-treated newborn calf serum instead of 10 % FCS. All other components remained unchanged. Assays for Dioxin inducibility of the integrated
30 DRE-TK-CAT construct were performed after a 48 hour treatment with Dioxin plus or minus resveratrol derivatives as described in the text. All chemicals applied to the cells were diluted in ethanol, control cells received ethanol alone. CAT expression was assayed on whole cell extracts (100 µg protein) with a CAT-ELISA assay (Roche, Meylan, France) according to supplier's specifications. Experiments were done in triplicate.

Ligand binding competition assay (Aryl Hydrocarbon Receptor)

5 Experiments were conducted exactly as previously described by Savouret et al., (3). Briefly, binding competition was performed using female New Zealand rabbit liver cytosol as the receptor source. The cytosol was prepared at 4°C in Hepes 20 mM pH 7.6, EDTA 1.5 mM, glycerol 10 %, β -mercapto-ethanol 10 mM, with Complete protease cocktail (Roche, Meylan, France) by homogenisation in a Ultra-Turax homogenizer (Bioblock Scientific, Illkirch, France) followed with 20 strokes in a Dounce Homogenizer with the tight pestle. The homogenate was centrifuged 30 min at 20,000 g. The supernatant was centrifuged at 105,000 g for 65 min. The cytosol was aliquoted, snap-frozen in liquid nitrogen and kept at -80 °C. Cytosol aliquotes (4.5 ml) were thawed on ice, diluted ten-fold in Hepes 20 mM pH 7.6, 5 mM CaCl₂, containing Complete protease cocktail and 10 mM mercapto-ethanol, and recentrifuged 30 min. at 105,000 g. The cleared cytosol dilution was adjusted to 0.85 mg proteins/ml. 930 μ l of diluted cytosol were preincubated with the desired amounts of unlabeled competitors in ethanol solutions for 1 hour at 4°C. Subsequently, 2,3,7,8-tetrachloro (1,6-³H) dibenzo-p-dioxin (28 Ci/mmol) was added at 0.2 nM and incubation continued for 3 hours at 4°C. Non-displaceable binding was assessed by incubating the aliquotes with 70 μ l of a 2% activated charcoal suspension in Hepes 20 mM pH 7.6 for 90 minutes at 4°C, followed by centrifugation at 15,000 g for 10 minutes. 500 μ l of the supernatants were counted in 5 ml of Ultima Gold cocktail (Packard, Meridien, CT) in a Beckman liquid scintillation Counter (45 % counting efficiency). Binding competition assays were repeated at least twice for each competitor and each point was performed in triplicate.

Ligand binding competition assay (Estrogen Receptor)

ER binding experiments were conducted using Estrogen Receptors extracted from MCF-7 cells as previously described (4). Briefly MCF-7 (from ATCC) were grown to 80% confluency in DMEM medium (Gibco BRL) supplemented with 10 % FCS. Cells were then scraped, and washed twice with PBS. After centrifugation for 10 minutes at 1500 rpm, the cells were resuspended in TM buffer (20 mM Tris, HCl pH 7.4; sodium molybdate, 20mM). Cells were broken by freeze-thaw lysis of the cell pellets in an equal volume of TM buffer. Cytosolic receptors were prepared by a 105,000 g x 60 minutes centrifugation at 0°C, and

then stored at -80°C. This cytosolic receptor solution was diluted to 60% in TM buffer, and then incubated with the corresponding ligand for 18 hr at 4°C in a volume of 100 µl with 50 µg of protein and 2 nM of [3H]Estradiol and two concentrations (1 and 10 µM) of unlabeled test ligand. Assays were terminated by loading 65 µl of the incubate on a 1.2 ml
 5 Sephadex™ LH-20 column equilibrated with the TM buffer. The flow through was collected and counted for radioactivity in ready safe scintillant (Beckman).

Cytotoxicity Assays

- 10 A MTT colorimetric assay was employed as described in (5) according to the established procedure 1. Drug stock solutions were prepared in EtOH and the final solvent concentration was = 1% EtOH(v/v), a concentration without effect on cell replication. Drugs were tested at 10 M on the human tumor cell line panel constituted of human lung carcinoma (A-549). Cells were cultured at 37°C in RPMI-1640 medium with streptomycin/penicillin and 5% (v/v)
 15 FCS in a humidified atmosphere containing 5% CO₂. Initial seeding densities varied among the cell lines to ensure a final absorbance reading in control cultures in the range of 0.7-1 A562 units. After the addition of the samples to the cell culture, the cells were incubated for 5 days before the MTT reagent was added. Each test was performed in duplicate and absorbance readings varied no more than 5%.

20

Biological effects of the stilbene derivatives of the invention

A. Dioxins and other AhR ligands

- 10 The biological effects of AhR ligands have been disclosed (ref. 1). Briefly, they are mediated by AhR is present in the cytosol of mammalian cells of almost all organs and tissues bound to heat shock protein 90 (hsp90). Upon binding to ligand, AhR dissociates from hsp90 and the ligand-AhR complex
 15 is translocated to the nucleus through association with a structurally related protein, the AhR nuclear translocator (Arnt). Inside the nucleus, the heteromeric ligand/AhR/Arnt complex regulates gene transcription by binding to DNA at dioxin-responsive enhancers (DRE) located within, or upstream
 20 of, a number of genes coding for phase 1 (cytochrome P-450) enzymes such as CYP1A1, 1A2, and 1B1 and phase 2 (detoxification) enzymes such as glutathione S-transferase Ya, aldehyde-3-dehydrogenase, and NAD(P)H:quinone oxidoreductase.

The most important phase 1 enzyme is CYP1A1 which is part of the aryl hydrocarbon hydrolase activity (AHH) and leads to the oxidative metabolism of AhR ligands, often to more pro-carcinogenic compounds. Phase 1 enzymes also increase the production of reactive oxygen (ROS) which have been shown to be associated with lipid peroxidation, oxidative DNA damage and other pathologic effects. AhR ligands also induce phase 2 enzymes responsible for detoxification and excretion of environmental toxicants.

Therefore, a complex activation of gene transcription occurs following exposure to AhR ligands, which may initiate toxic effects as well as attempts to clear the ligand or its active metabolites from the body.

The prototype AhR ligand is 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD or dioxin) which binds to the AhR with the highest affinity of all known compounds ($K_d = 10^{-10}$ to 10^{-11} M in mice and 10^{-9} M in humans).

It results from the inventor's demonstration of the AhR inhibitory activity of stilbene derivatives that a variety of AhR ligand-induced genes can be repressed. This directs the uses of stilbene derivatives into a number of different areas. By searching gene databanks (such as EMBL and Genbank) for genes containing one or several putative dioxin responsive elements (DRE) presenting the consensus sequence TNGCGTG, or GNGCGTG in any orientation, preferentially in their promoter region.

Outside of the already known phase 1 and phase 2 enzymes, it was discovered that a number of genes contain DRE's and, thus are likely to be regulated by AhR ligands and resveratrol. A number of these genes are listed in two tables according to their relationship to distinct physiological or pathological events.

Table 3 lists various genes involved in hormonal control, lipid metabolism and inflammation, genetic or degenerative diseases, proliferative diseases and cancers.

Table 4 lists human viruses and related human proteins.

Table 3: Human genes containing DRE's in their regulatory regions. The bovine prion gene is also shown. Cds: coding sequence.

GENES	Access Nr	Sites	Location	Comments
Enzymes				
inducible NOS	X97821	4	promoter	
endothelial NOS	U24214	1	promoter	
Cyclooxygenase 2	U20548	1	promoter	
PG H Synthase	U44805	1	promoter	
PG endoperoxide synthase	M31812 D64068	1	promoter	
5-Lipoxygenase	M38191	5	promoter, cds	
15-lipoxygenase	U63384	6	promoter, cds	
Phospholipase A2	U11239	2	promoter	
Thromboxane A2 receptor	D15054	1	promoter	
Disease-related proteins				
ICAM (CD54)	M65001	promoter	2	
Adenomatous polyposis protein	U02509	2	promoter, cds	
Adenomatous polyposis protein	D15951	1	supplementary exon in 5'UTR	non-coding, brain specific
Adenomatous polyposis protein	D15980	1	5' UTR	multiple variants
Human Prion protein	D00015	2	cds	Creutzfeld-Jacob
[Bovine Prion protein]	D26150	4	intron 1	bovine gene
Ob (leptin)	U43589	5	promoter, cds	obesity
presenilin	L76518	5	promoter	Alzheimer
STM2	U50871	7	promoter	Familial Alzheimer
GADD p153	S40707	2	promoter	DNA damage
Oncogenes				
WT 1b	S77896	1	promoter	Wilm's tumor
WNT-5A	U39837	3	promoter	
PCNA	J05614	1	promoter	
p53	J04238	17	complete gene	apoptosis, cell cycle
p53	J04238	1	promoter	
c-Ha-Ras	M13221	1	promoter	
c-jun	U60581	3	promoter	
c-myc p64	M13930	4	promoter	
c-raf-1	M38134	2	promoter	
ATF 3	U37542	2	promoter	

Table 4: Viral genes or genomes containing DRE's. UTR: untranslated region. LTR: long terminal repeat. IE: immediate early. Cds: coding sequence.

VIRAL GENE	ACCESS No	LOCATION	SITES Nr	COMMENT
Cytomegalovirus IE-1	M64941	promoter	1	
HTLV-1	S76263	5' LTR	1	
HTLV-1	S76263	TAX gene	1	
HTLV-4	X06392	5' LTR	2	
HIV 1 isolate BRU (LAV-1)	K02013	3' LTR	1	repeatedly found in all isolates
HIV 2	U22047	genome	2	
HIV2 EHO	U27200	genome	1	
CD4	U01066	promoter	1	HIV receptor
CXCR4	AF005053	promoter, cds	1	HIV co-receptor
Human Papilloma Virus (HPV)	X52061	genome for	1	long control region
HPV 11	J04351	genome	2	
HPV 16	M33616	genome	1	3' flanking region
HPV 25	D50264	promoter	1	gene E6
HPV 33, 47, 58 59		genome	1	
Human poliovirus 1	X70509	5' UTR	1	
Human rhinovirus 18	D00239	genome	2	
Human rhinovirus 2, 14, 89		genome	1	
Herpes virus HSV1	U18080	genome	1	IE transactivator gene
Herpes virus HSV1	M13885	genome	1	"a" segment
Hepatitis A virus	K02990	genome	1	
Hepatitis B virus	X98077	genome	1	
Hepatitis C virus	D30613	genome	4	
Adenovirus type 2	J01917	genome	21	
Adenovirus type 12	J01910	genome	1	right terminal repeat
Adenovirus type 12	X14757	promoter	1	E1A gene
Adenovirus type 19A	X95259	early region 3	1	HLA binding protein gene
Adenovirus type 41	M18289	genome	1	E1A gene
Human respiratory syncytial virus	M17212	cds	1	attachment protein gene
Human influenza virus all registered subtypes		genome: multiple sites in various genes		M1, M2 proteins matrix proteins, hemagglutinin, neuraminidase,

B. Cardiovascular effects of AhR ligands

Cigarette smoke contains benzo[a]pyrene (BaP) and other AhR ligands such as polychlorinated dibenzo-p-dioxins (PCDD) in the tar fraction. It has been estimated that the amount of BaP in mainstream cigarette smoke is between 40 and 100 ng per cigarette and that up to 460 ng of BaP per hour can be inhaled by non-smokers in a smoking environment. Daily intake of PCDDs by smoking 20 cigarettes a day has been estimated to be 4.3 pg/kg body weight/day. In heavy smokers, daily exposure to BaP could be as high as 0.05 mg/kg in addition to other unquantifiable AhR ligand exposure from cigarettes and from the environment. The AhR ligands in cigarette smoke are present in high enough concentrations to induce CYP1A1 and AHH activity in the lungs, placenta, kidney, and ovary and in the endothelial cells lining blood vessels. AHH metabolism of BaP results in reactive carcinogenic intermediates which bind to DNA forming predominantly covalent adducts. Cigarette smoking has been identified as the direct cause of at least eight different human cancers including lung, bladder, gastrointestinal tract, and leukemia, as well as ischemic heart disease. There is also epidemiologic evidence linking smoking with acceleration of acquired immune deficiency (AIDS) and with osteoporosis and increased risk of fractures in both men and women.

In the cardiovascular system, AhR ligands may be atherogenic by disrupting the functions of endothelial cells in blood vessels. AhR ligands have been shown to induce CYP1A1/AHH in porcine aortic endothelial cells with ED50 values of 180 nM for BaP and 0.015 nM for TCDD. Stimulation of these phase I enzymes leads to increased ROS production which may cause endothelial cell membrane damage and lipid oxidation. Oxidized LDL formed by this mechanism undergoes unregulated uptake by a scavenger receptor in vascular endothelial cells leading to over-accumulation and the formation of "foam

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cells". It has also been demonstrated that AhR ligands can disrupt endothelial integrity and permeability, which could allow increased uptake of cholesterol-rich lipoprotein remnants into the arterial wall leading to atheroma formation. This effect was only seen with PCBs that bind to the AhR and induce CYP1A1. In addition, the same PCBs increased oxidative stress and lipid peroxidation in cultured endothelial cells, which may be the mechanism of cell injury, and which was correlated in time with increases in CYP1A1 induction.

C. Activation of the progression of HIV and other viral infection by AhR ligands

AhR ligands are known to enhance the replication of viruses in animals.

It is believed that activation of latent human immunodeficiency virus type 1 (HIV-1) expression by AhR ligands, is important for the progression of acquired immune deficiency disease (AIDS). It has been shown that dioxin and other AhR ligands activate HIV-1 expression by induction of CYP1A1 _____ and by activation of the cellular transcription factor, nuclear factor kappa-B (NF-kappa B) _____

— The genes for both of these factors (CYP1A1, NF-kappa B), as well as the long terminal repeat (LTR) of the HIV gene promoter itself, contain DRE's consistent with their modulation by AhR ligands. Moreover, the two known entryways of HIV into lymphocytes, the CXCR4 chemokine receptor and the CD4 glycoprotein also present multiple DRE's in their promoters (see table 4)

D. Skin disorders associated with dioxins exposure

AhR ligands, especially dioxins, are potent inducers of chloracne. In every industrial accident involving these environmental toxins, _____ chloracne was considered the hallmark of dioxin intoxication _____ and was consistently observed in the majority of exposed individuals. This acneiform eruptive skin condition occurs as a result of an altered pattern of differentiation of the basal cells of sebaceous glands and an altered rate of differentiation of keratinocytes, _____. These cells express increasing concentrations of AhR during ongoing differentiation _____ and, moreover, AhR ligands themselves enhance keratinocyte terminal differentiation _____. It is likely that AhR

ligands elicit specific skin responses through activation of the AhR resulting in modified levels of gene expression in the skin.

E Osteoporosis and exposure to AhR ligands

There are epidemiologic data to support cigarette smoking as a risk factor for osteoporosis in both men and women. _____ The plausibility of this relationship is strengthened by the dose-response relationship between the number of cigarettes smoked per day and the decrease in bone mineral density _____, and by the association between smoking and increased fracture risk _____. In addition, studies of twins discordant for cigarette smoking have demonstrated that bone density of women who smoked, or who smoked more heavily, was found to be significantly lower than that of their twin sisters. _____ The pairs with the largest difference in smoking had the largest difference in bone density. The mechanism by which smoking and osteoporosis are related remains unknown. However, it is unlikely to be a simple direct effect since the relationship of ovarian steroids and bone turnover is so important.

It is believed by the inventors that AhR ligands present in cigarette smoke, and in the environment, are responsible for loss of bone density through 3 different interconnected effects 1) An anti-estrogenic effect of AhR ligands in bone since AhR ligands have been shown to inhibit a broad spectrum of estrogen-induced responses in rodents and in breast cancer cell lines 2). A direct toxic effect of AhR ligands on bone. Both AhR and Arnt have been found to be expressed in bone tissue of mice during embryonic development. The effect of TCDD was examined on the organization of bone tissue in vitro using primary cultures of normal diploid calvarial-derived rat osteoblasts. It was found that TCDD inhibited formation of bone tissue-like multicellular nodules suggesting a direct association with reduced bone mass. 3). A direct toxic effect of AhR ligands on the ovary. There is compelling evidence for a direct toxic effect of AhR ligands on the ovary. Women who smoke undergo menopause at an earlier age (between 1 and 4 years earlier) than non-smokers. In addition, the incidence of ovarian cancer in women is highest in industrialized urban areas and in women working in rubber, electrical or textile industries where exposure to AhR ligands are increased compared to other industries. Experimental data in animals showing direct toxic effects of BaP and other AhR ligands on oocytes support these epidemiological studies.

F. Resveratrol, the natural molecule.

The low rate of coronary heart disease in France compared to other western countries, despite the presence of similar risk factors (high animal fat intake, low exercise levels, and high rate of smoking) has been called the French paradox _____. However, France was found to have the highest consumption of wine of any of the countries studied, leading to the speculation that wine may contain cardioprotective compounds. In addition to ethanol, which in moderate consumption appears to reduce mortality from coronary heart disease by increasing high-density lipoprotein cholesterol and inhibiting platelet aggregation.

Two flavonoids (catechin, quercetin) and the tri-hydroxystilbene, resveratrol, inhibit the synthesis of thromboxane in platelets and leukotriene in neutrophils, and modulate the synthesis and secretion of lipoproteins in animals and in human cell lines.

_____ However, flavonoids should be avoided in the treatment of human diseases due to their numerous harmful side-effects.

Resveratrol (3,5,4'-trihydroxystilbene) is the parent compound of a family of molecules, including glucosides and polymers, existing in cis and trans configurations in a variety of plants classified as spermatophytes of which vines, peanuts and pines are the prime representatives. _____ Resveratrol is produced by plants as a phytoalexin or antifungal agent. Resveratrol is usually present in red wine in concentrations between 1 and 8 mg/L (trans-resveratrol: 1 to 5 mg/L; and cis-resveratrol: 0.5 to 4 mg/L).

Another recent study demonstrated that resveratrol prevented chemical induction of pre-neoplastic lesions in a mouse mammary gland culture model and could slow down the growth of skin tumors which had been initiated in mice by a two step carcinogenic stimulus. This effect was proposed to act through the inhibition of cyclo-oxygenase and hydroperoxidase enzymes, by induction of phase 2 drug-metabolising enzymes, by anti-oxidant activity and by inducing differentiation of cancer cells.

Since resveratrol is highly lipophilic, it is poorly absorbed from fruit and vegetables in the diet. However, it is readily soluble in alcohol, and is present in therapeutic levels in many red

wines which prove to be a good delivery system. This fact may be an advantage for wine drinkers such as the majority of the adult French population, but means that abstainers and small children are not exposed to the protective effects of resveratrol against AhR ligands. In addition, since dioxin and other hydrophobic AhR ligands are also soluble in alcohol, an alcoholic delivery system will increase the absorption of the very toxins one is trying to antagonize. It is preferable for resveratrol to be administrated as a dietary supplement, dissolved in a small volume of oil or other innocuous lipophilic solvent.

The discovery by the inventors of the potent anti-dioxin effects of resveratrol, has important medical and socioeconomic implications. It has been demonstrated that resveratrol, in concentrations easily obtained clinically, can inhibit dioxin-induced phase I enzymes activity as well as interleukin-1 beta production and HIV promoter induction. It can therefore protect against a variety of diseases and toxic effects related to exposure to AhR ligands.

The obtained data show that the known AhR associations with immunosuppression, cancer and viral infections such as HIV-induced AIDS may be prevented by AhR antagonists such as resveratrol. In addition, the constant presence of DRE's in inflammatory related genes such as IgE, cytokines, chemokines, their receptors and the iNOS gene suggests a role of AhR and its ligands in inflammatory diseases. Examples of such iNOS-mediated diseases include atopic dermatitis, rheumatoid and osteo-arthritis, neurodegenerative diseases (multiple sclerosis, amyotrophic, lateral sclerosis), diabetes, recurrent abortion, and likely, several others. The presence of DRE's in genes for chemokines and their receptors further supports the link between AhR ligands and inflammation and fever.

Finally, the presence of DREs in the intronic enhancer of the bovine Prion gene (responsible for bovine spongiformis encephalitis and highly suspected to be involved in the new variant of human Creutzfeld-Jacob's disease) and in the promoter region of presenilin and STM2 genes (Alzheimer's disease) suggests that AhR ligands may aggravate these

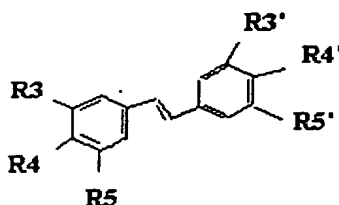
5 neurodegenerative conditions.

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CLAIMS

1. Stilbene derivatives having formula I



wherein

R3, R4 and R5 and R3', R4' and R5' are identical or different and represent H, OH, O-alkoxy or hal, said alkoxy group being a C1-C6 alkoxy and "hal" being F, Cl or CF₃, with the proviso that one of R4', R3 and R5 or R4, R3' and R5' does not represent OH, OCH₃ or OCH₂CH₃ when the two other substituents are both OH, OCH₃, OCH₂CH₃, respectively,

and the symmetrical derivatives.

2. The trans isomers of the stilbene derivatives of claim 1.

3. The trans isomers of claim 2, wherein R3 and R5 are hal.

4. The trans isomers of claim 3, wherein R3' or R4' is hal, alkoxy or hydroxy, and R5' is H.

5. The trans isomers of claim 3 or 4, wherein R3 and R5 are Cl.

6. The trans isomers of claim 5, wherein R3' is H, and R4' or R5' is Cl or methoxy.

7. The trans isomers of claim 2 selected in the group comprising (E)-1-(4'-trifluoromethylphenyl)-2-(3,5-ditrifluoromethylphenyl)-ethene, (E)-1-(4'-methoxyphenyl)-2-(3,5-dichlorophenyl)-ethene, and (E)-1-(4'-chlorophenyl)-2-(3,5-dichlorophenyl)-ethene which bound to AhR with respective relative binding affinity of 52.1, 112.0 and 130.0, without detectable affinity for ER.

8. The symmetrical derivatives of the trans isomers of anyone of claims 3 to 7, particularly the derivatives with R3', R5' and R4 or R3', R5' are hal. and R4 is OH or alkoxy.

5 9. The use of the stilbene derivatives according to anyone of claims 1 to 8 as antagonists of AhR ligands.

10. Pharmaceutical compositions comprising an effective amount of at least one stilbene derivative according to anyone of claims 1 to 8, with a pharmaceutically acceptable carrier.

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11. The pharmaceutically compositions of claim 10 in a form for administration by the oral, nasal, parenteral or topical route.

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12. The pharmaceutical compositions of claim 11, wherein said form is a gel, capsules, drops, syrup or alcohol syrup, for administration by the oral route, spray or drops for administration by the nasal route, solution for administration by the parenteral route, and cream, ointment, shampoo or lotion for application by the topical route, the vehicle comprising an oil or a pharmaceutically acceptable alcohol.

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13. The pharmaceutical compositions of anyone of claims 10 to 12, said comprising then administration at a dosage from 0.1 mg to 5 g/day, especially from 20 to 200 mg/day and in particular from 10 to 100 mg/day.

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14. The pharmaceutical compositions of anyone of claims 10 to 13 for the treatment of dermatitis, acne, psoriasis, hyperkeratotic lesions, eczema, or skin aging and wrinkling associated with common environmental exposure to AhR ligands.

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15. The pharmaceutical compositions of anyone of claims 10 to 13 for preventing or avoiding the development of cold or flu symptoms related to viral infections aggravated by AhR ligands.

16. The pharmaceutical compositions of anyone of claims 10 to 13 for the prevention of AhR ligand-induced triggering of HIV (and other viruses) gene expression and progression of AIDS, particularly for the treatment of viral infections such as HIV-induced AIDS.

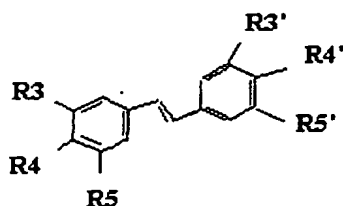
17. The pharmaceutical compositions of anyone of claims 10 to 13 for the prevention of prion-induced Spongiformis Encephalitis in humans and livestock.
- 5 18. The pharmaceutical compositions of anyone of claims 10 to 13 for the prevention of osteoporosis in reproductive age women and for the prevention and treatment of osteoporosis, alone or either in association with hormone replacement therapy or calcium and vitamin D in post-menopausal and elderly women.
- 10 19. The pharmaceutical compositions of anyone of claims 10 to 13 of inflammatory conditions caused by excessive nitric oxide and/or immunoglobulin E production such as: atopic dermatitis, rheumatoid and osteo-arthritis, neurodegenerative diseases (such as Alzheimer, multiple sclerosis, amyotrophic lateral sclerosis), diabetes.
- 15 20. The pharmaceutical compositions of anyone of claims 10 to 13 for reduction of fever associated with bacterial, viral, or allergic illnesses.
- 20 21. The pharmaceutical compositions of anyone of claims 10 to 13 for the treatment of obstetrical and gynecologic conditions such as endometriosis, fibroids (leiomyoma), pre-eclampsia and recurrent abortion.
- 25 22. Use of the stilbene derivatives according to anyone of claims 1 to 8, as food additive in powdered or liquid formula, cereal, and in canned food to prevent the toxic effects of environmental exposure to AhR ligands.
23. Use of the stilbene derivatives according to anyone of claims 1 to 8, for impregnating cigarette filters.

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**“NEW STILBENE DERIVATIVES AND THEIR USE AS ARYL HYDROCARBON
RECEPTOR ANTAGONISTIC LIGANDS”**

ABSTRACT

The invention relate to stilbene derivatives having formula I



wherein

R3, R4 and R5 and R3', R4' and R5' are identical or different and represent H, OH, O-alkoxy or hal, said alkoxy group being a C1-C6 alkoxy and “hal” being F, Cl or CF₃, with the proviso that one of R4', R3 and R5 or R4, R3' and R5' does not represent OH, OCH₃ or OCH₂CH₃, when the two other substituents are both OH, OCH₃, OCH₂CH₃, respectively,

and the symmetrical derivatives.

Use of said stilbene derivatives particularly as pharmaceutical compositions and food additives.

No drawings

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